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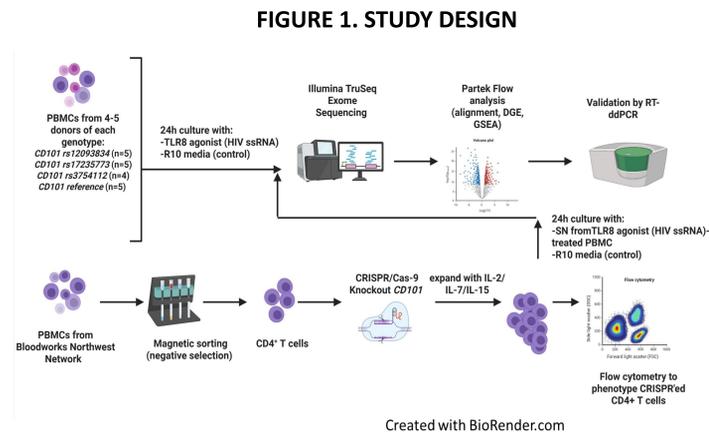
BACKGROUND

- Genetic variants in the gene **CD101** increase HIV risk (hazard ratio [HR]=4.3, $p=1.9e^{-4}$)¹ while **AXL** variants reduce the risk (HR=0.4, $p=4.7e^{-4}$)². Nevertheless, the immunological consequences of **CD101** and **AXL** variants remain unclear at the cellular and molecular levels.
- CD101** is a transmembrane regulator of T cell activation expressed on multiple immune cells. In T cells, it inhibits IL-2 secretion, activation, and proliferation³.
- AXL**, a receptor tyrosine kinase, inhibits toll-like receptor (TLR)-mediated innate responses by blocking interferon-stimulated genes (ISGs) in monocytes, resulting in inhibition of the inflammasome⁴.
- We previously showed that peripheral blood mononuclear cells (PBMC) from individuals with **CD101** variant alleles exposed to a **TLR8 agonist** (HIV single-stranded RNA, HIV ssRNA) showed a larger reduction of **CD101** than in PBMC bearing reference **CD101** alleles⁵.
- Our goal was to understand how **CD101** variants increase HIV-1 susceptibility compared to the reference.

METHODS (FIGURE 1)

- Using exome-targeted RNA-Seq, we evaluated transcriptomic profiles in PBMCs from individuals bearing each of the five missense variants (*rs3754112*, *rs17235773*, *rs116063197*, *rs12093834*, *rs34882009*) associated with increased HIV acquisition risk. We compared them to those from PBMCs bearing reference **CD101** alleles.
- We also used CRISPR to knock out **CD101** in primary CD4⁺ T cells to test its impact on transcriptomics after challenge with HIV ssRNA.
- We compared: 1) Unstimulated PBMC from individuals with reference vs variant **CD101** genotypes (n=5 and 14, respectively), 2) **CD101** reference vs variant PBMC stimulated with HIV ssRNA – a TLR8 agonist, 3) primary CD4⁺ T cells with **CD101** (**CD101**^{+/+}) vs with **CD101** knocked out (**CD101**^{ko/ko}) via CRISPR and 4) **CD101**^{+/+} vs **CD101**^{ko/ko} primary CD4⁺ T cells each exposed to supernatants from HIV ssRNA-treated PBMC.
- Sequence alignment, differential gene expression (DGE), and gene set enrichment analyses were done using the Illumina Partek Flow pipeline.
- AXL** and **CXCR4** gene expression validation was done using digital droplet PCR (ddPCR).

This study reveals that **CD101** and **AXL** are expressed in an HIV RNA-sensing inflammatory pathway with risk-associated **CD101** variants linked to both stronger **CD101** downregulation and increased **AXL** induction.



RESULTS

FIGURE 2. EXPOSURE OF PBMC TO HIV ssRNA, A TLR8 AGONIST, UPREGULATES PROINFLAMMATORY GENES AND INTERFERONS

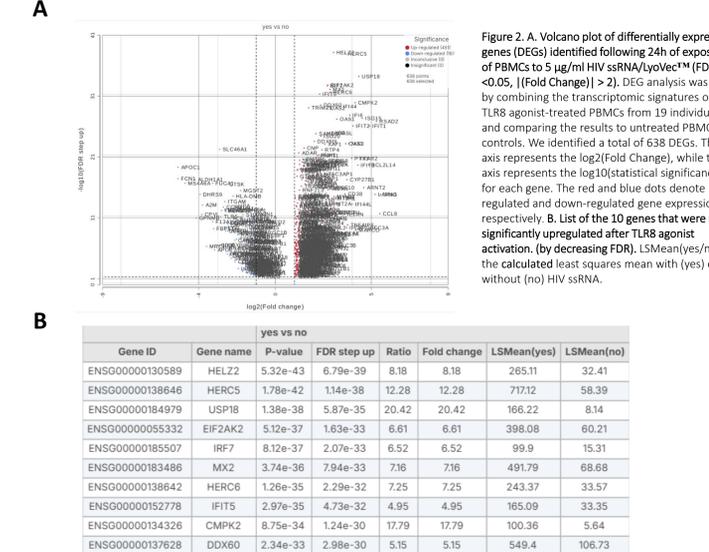


FIGURE 3. CD101 mRNA EXPRESSION WAS DOWNREGULATED FOLLOWING HIV ssRNA STIMULATION, WHEREAS THE AXL RESPONSE WAS VARIABLE

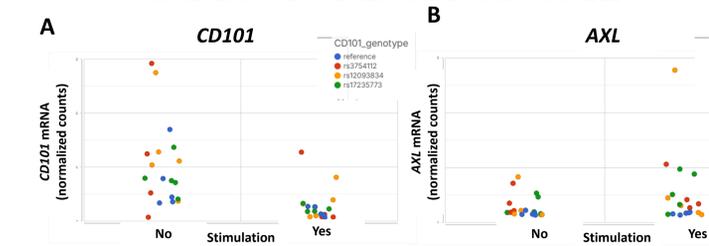
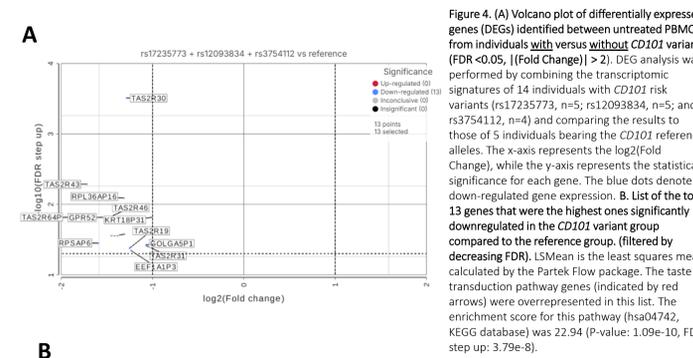


Figure 3. A. Dot plots depicting the level of expression of **CD101** (A) and **AXL** (B). TMM (trimmed mean of M-values)-normalized read counts for each gene are represented on the Y-axis. A point represents each sample; samples are separated by treatment (No=untreated control; Yes=24h HIV ssRNA stimulation) and by **CD101** genotype, identified by colors, as shown in the legend.

FIGURE 4. IN THE UNSTIMULATED CONDITION, GENES RELATED TO TASTE TRANSDUCTION WERE DOWNREGULATED IN THE CD101 VARIANT GROUP COMPARED TO THE REFERENCE GROUP



Gene ID	Gene name	P-value	FDR step up	Ratio	Fold change	LSMean(rs17235773 + rs12093834 + rs3754112 vs reference)	LSMean(reference)
ENS000000256188	TAS2R30	2.4e-8	0.00031	0.41	-2.43	7.95	19.35
ENS000000255374	TAS2R43	8.06e-7	0.0052	0.3	-3.29	2.82	9.29
ENS000000232228	RPL36AP18	0.0000025	0.0081	0.4	-2.49	1.9	4.72
ENS000000249850	KRT18P31	0.0000072	0.015	0.5	-2.01	4.06	8.15
ENS000000203737	GPR52	0.0000086	0.015	0.34	-2.92	2.08	6.09
ENS000000256274	TAS2R64P	0.000011	0.015	0.26	-3.78	0.83	3.12
ENS000000226761	TAS2R46	0.000012	0.015	0.36	-2.78	1.92	5.34
ENS000000258111	---	0.000024	0.027	0.4	-2.48	1.67	4.14
ENS000000214629	RPSAP6	0.000036	0.036	0.33	-3.02	1.45	4.38
ENS000000251215	GOLGASP1	0.000041	0.037	0.48	-2.1	2.07	4.36
ENS000000256436	TAS2R31	0.000045	0.039	0.48	-2.09	5.16	10.8
ENS000000212124	TAS2R19	0.000055	0.042	0.42	-2.38	3.27	7.78

FIGURE 5. FOLLOWING HIV ssRNA STIMULATION, SIX GENES WERE DIFFERENTIALLY EXPRESSED BETWEEN THE CD101 VARIANT GROUP AND THE REFERENCE

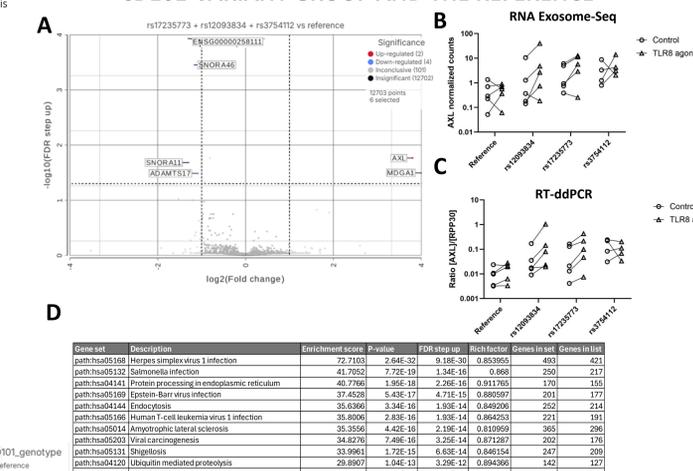


Figure 5. (A) Volcano plot of differentially expressed genes (DEGs) identified between TLR8-activated PBMC from individuals with versus without **CD101** variants (FDR <0.05, [Fold Change] > 2). DEG analysis was performed by combining the transcriptomic signatures of 14 individuals with **CD101** risk variants (*rs17235773*, *n=5*; and *rs3754112*, *n=4*) and comparing the results with those from 5 individuals bearing the **CD101** reference alleles. The x-axis represents the log2(Fold Change), while the y-axis represents the statistical significance for each gene. The red and blue dots denote up-regulated and down-regulated gene expression, respectively. (B) **AXL** counts determined by RNA-exosome Seq. Normalized values are shown on the y-axis; **CD101** genotypes are indicated on the x-axis. (C) Validation of RNA-exosome results by RT-digital droplet PCR (RT-ddPCR). The y-axis denotes the normalized copies of **AXL** (using *RPP30* counts as a housekeeper). (D) Pathway enrichment analysis (KEGG datasets), filtered by decreasing FDR step up. The pathways that were overrepresented in the variant groups are related to intracellular defense and protein folding.

ACKNOWLEDGEMENTS

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FIGURE 6. COMPARISON OF TRANSCRIPTOMICS BETWEEN CD101+/+ AND CD101ko/ko CD4+ T CELLS

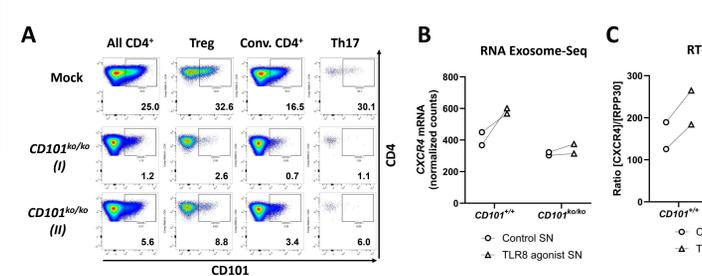


Figure 6. (A) Confirmation of **CD101** gene ablation by flow cytometry. **CD101** CRISPR-edited CD4⁺ T cells were expanded in R10 media supplemented with IL-2, IL-7, and IL-15 and analyzed by flow cytometry. **CD101** expression was measured at the cell surface along with lineage markers. Mock control was done by performing the nucleofection reaction without **CD101** sgRNAs. Two independent editing experiments ("I" and "II") were done on CD4⁺ T cells from the same donor. "All-CD4"⁺=CD45⁺CD3⁺CD4⁺; "Conv. CD4"⁺=CD45⁺CD3⁺CD4⁺CD25⁺CD127⁺; "Treg"⁺=CD45⁺CD3⁺CD4⁺CD25⁺CD127⁺FoxP3⁺; "Th17"⁺=CD45⁺CD3⁺CD4⁺CD25⁺CD127⁺CCR6⁺CD161⁺. (B) **CXCR4** counts determined by RNA-exosome Seq. Normalized values are shown on the y-axis; **CD101** genotypes are indicated on the x-axis. (C) Validation of RNA-exosome results by RT-digital droplet PCR (RT-ddPCR). The y-axis denotes normalized **CXCR4** copies (using *RPP30* counts as a housekeeper).

CONCLUSIONS

- Exposure of PBMCs to HIV ssRNA resulted in a strong induction of proinflammatory and antiviral genes (Fig. 2), with concomitant downregulation of **CD101**, whereas **AXL** response was variable (Fig. 3).
- In unstimulated PBMCs, 13 genes were downregulated in the **CD101** variant compared to the reference group, with an overrepresentation of genes related to taste transduction (Fig. 4)
- Following TLR8 agonist stimulation, 6 genes (**AXL**, **SNORA46**, **SNORA11**, **MDGA1**, **ADAMTS17**, and **ATP5G1**; Fig. 5) were differentially expressed between variant and reference. Remarkably, **AXL** was upregulated 14-fold in PBMCs with **CD101** variants relative to the reference (FDR=0.02).
- Comparison of reference vs variant cells in both the stimulated and unstimulated conditions revealed overrepresentation of pathways related to protein processing and intracellular defense (Fig.5).
- Comparing **CD101**^{+/+} to **CD101**^{ko/ko} CD4⁺ cells resulted in no DEGs. However, when the **CD101**^{ko/ko} and **CD101**^{+/+} cells were exposed to supernatants from the stimulated PBMC, **CXCR4** was down-regulated in the **CD101**^{ko/ko} cells (1.7-fold change, $p=1.67E-6$, Fig.6).

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PLAIN LANGUAGE SUMMARY

People who carry certain natural changes in the immune gene **CD101** show altered activity of other immune genes, including higher levels of the interferon response repressor **AXL**, which together may make it easier for HIV to establish infection. By removing the function of **CD101** in immune cells, we also observed changes in the HIV coreceptor **CXCR4**, suggesting that **CD101** reshapes inflammatory pathways in ways that can influence a person's vulnerability to HIV.